CROSS-TALK BETWEEN MAPKs AND PI-3K PATHWAYS ALTERS THE FUNCTIONAL DENSITY OF $I_K$ CHANNELS IN HYPERTROPHIED HEARTS

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INTRODUCTION

Cardiac hypertrophy is an adaptive response to the inciting stress produced by workload to the heart. Chronic volume-overload occurring during aortic insufficiency, mitral regurgitation or arteriovenous shunt leads to the development of eccentric cardiac hypertrophy. It is characterized by ventricular chamber enlargement disproportionate to ventricular wall thickening. This involves the action of specific growth factors which are activated during the development of cardiac hypertrophy. Insulin-like growth factor-1 (IGF-1) has been known for its major role in cellular proliferation and differentiation during heart development. Furthermore, in the volume-overload model, we have shown that cardiac myofibers have an increased receptor affinity to IGF-1, which corroborated with the induction of eccentric hypertrophy. This was confirmed by our recent findings on the major role that IGF-1-dependent intrinsic signaling pathways, MAPK and PI-3 kinase, play in modulating myocardial electrophysiological function.

Binding of IGF-1 to its receptor stimulates PI-3 kinase causing activation of Akt. Transgenic overexpression of Akt was sufficient to induce significant cardiac hypertrophy in mice without affecting systolic function. Furthermore, IGF-1 activated the mitogen-activated protein kinase (MAPK) cascade in addition to the PI-3K/Akt one. Lavender et al. showed that short exposure of ventricular cardiomyocytes to IGF-1 leads to the development of cardiac hypertrophy through activation of downstream ERK-MAPK pathway. In addition, Shyu et al. found that the increased expression of myostatin in hypertrophied cells is mediated by IGF-1, partly through p38 MAP kinase. In the same line of evidence, we have recently shown that IGF-1 plays an important role in the development of eccentric cardiac hypertrophy, through the activation of MAPK and PI-3K pathways.

However, there is still a lack of information regarding the interactions between the MAPKs and the PI-3K pathways in response to IGF-1 in vitro or in vivo. Therefore, the objective of this study was to determine the level of interaction between the MAPK (ERK1/2 and p38) as well as the PI-3K in mediating IGF-1 effects in the normal and hypertrophied hearts. We hypothesized that during the compensated phase of eccentric cardiac hypertrophy, IGF-1 activation of MAPKs and PI-3 kinase induces differential cross-talk between these kinases that can be a key element in modulating its effect through the progression of the disease.

MATERIALS AND METHODS

All procedures used conform to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH) publication No. 85-23, revised 1996.
Eccentric Cardiac Hypertrophy

Induction of Volume-overload Cardiac Hypertrophy

Adult male Sprague-Dawley rats (200–250 g) were used. The abdominal aorta and vena cava were shunted at the union of the segment two-third caudal to the renal artery and the one-third cephalic to the aortic bifurcation with an 18G needle. The entire was hung on Langendorff apparatus for 15 minutes with Tyrode’s solution containing either 1) regular Tyrode’s solution or 2) IGF-1, or 3) Tyrode’s solution containing specific kinase inhibitor followed by IGF-1. At the end of the experiment, hearts were quickly snap-frozen in liquid nitrogen for western blot processing.

Isolated Heart Retrograde Perfusion

Immediately after heart excision, it was hung on Langendorff apparatus for 15 minutes with Tyrode’s solution containing either 1) regular Tyrode’s solution or 2) IGF-1, or 3) Tyrode’s solution containing specific kinase inhibitor followed by IGF-1. At the end of the experiment, hearts were quickly snap-frozen in liquid nitrogen for western blot processing.

Western Blotting

Activation of ERK1/2, p38 MAPK and Akt were assessed using western blot technique as previously described. Bands were visualized by Chemiluminescence. Films from at least three independent experiments were scanned and densities of the immunoreactive bands were evaluated using UnScanIT software. Kinase activities were evaluated as the ratio of phosphorlated kinase over total protein kinase per experiment.

Isolation of Adult Rat Cardiomyocytes

As previously described, retrograde perfusion of the coronary arteries was performed by Langendorff method. The heart was sequentially perfused with Tyrode’s solution containing lliberase Blenzyme 4 (Sigma, Missouri). Then the heart was perfused with regular Tyrode’s solution. Finally, the ventricles were cut, minced and filtered. The cells were diluted and kept in Tyrode’s solution until experimentation.

Electrophysiological Studies

The whole-cell patch-clamp technique was used in the voltage-clamp mode. Ventricular myocytes were perfused with a buffer containing (mM): 130 K-glutamate, 20 KCl, 5 EGTA, 5 NaCl, 5 Mg-ATP, 1 MgCl₂, 10 HEPES, and pH at 7.2. The current amplitudes were plotted as a function of the command potential. The inhibitors for PI-3K (LY294002), ERK1/2 MAPK (PD98059) and p38 MAPK (SB203580) were purchased from Cell Signaling Technology, Mass.). Bands were visualized by Chemiluminescence. Films from at least three independent experiments were scanned and densities of the immunoreactive bands were evaluated using UnScanIT software. Kinase activities were evaluated as the ratio of phosphorlated kinase over total protein kinase per experiment.

Experimental Protocols for Potassium Currents

Step voltages (400 msec) in 10 mV increments between -40 mV and +40 mV were applied from a holding potential of -40 mV. Steady-state currents were plotted as a function of the command potential. The inhibitors for PI-3K (LY294002), ERK1/2 MAPK (PD98059) and p38 MAPK (SB203580) were purchased from Cell Signaling Technology, Mass.).

Statistical Analysis

All statistical analysis was performed using SigmaStat software. The paired Student’s t-test was used to compare data before and after drug treatment of the same animal group. The heteroscedastic two-sample unpaired Student’s t-test assuming unequal variances was used when comparing the drug effects between two different animal groups (sham vs shunt). In addition, we performed one-way ANOVA followed by ad hoc Dunnett’s test for comparing the differences between the effects of inhibitors to a control in each animal model (sham or shunt) and on each kinase. Using the null hypothesis, P≤.05 was considered significant.

RESULTS

Structural Parameters

The data on the structural parameters from sham and shunted adult rats confirmed the development of the eccentric cardiac hypertrophy within 3 weeks post-surgery, as seen in Table 1. The shunted rats did not show a significant difference in

| Table 1. Differences in the structural parameters of sham and shunt hearts |
|---------------------------------|---------------------------------|---------------------------------|---------------------------------|
| **Body Weight (g)**            | **Heart Weight (mg)**           | **Relative Heart Weight (mg/100g body weight)** | **Membrane Capacitance (pF)** |
| Sham 225 ± 12                  | 1043 ± 22                      | 474 ± 19                        | 209 ± 14                       |
| Shunt 256 ± 21                 | 1397 ± 77*                     | 674 ± 34*                       | 294 ± 24*                      |

* n=12 for each group
* P<.01 for sham versus shunt.
their body weight vs normal sham ones. However, the heart weights ($P<.01$), as well as the relative heart weight of shunted rats, were higher compared to the sham ones ($P<.01$). In addition, the cellular membrane capacitance was greater in the cardiomyocytes from hypertrophied hearts as compared to control ones ($P<.01$).

Effects of IGF-1 on ERK1/2 and p38-MAPKs as well as Akt in Normal and Hypertrophied Hearts

The activation levels of ERK1/2, p38 MAPKS and Akt was higher in the
hypertrophied cardiomyocytes compared to normal ones, as shown in Figure 1A. Cardiac hypertrophy activated ERK-1 by 56 ± 19% ($P=0.05$), ERK-2 by 227 ± 21% ($P<.001$), p38 MAPK by 75 ± 17% ($P=.01$) and Akt by 67 ± 19% ($P=.02$). Similarly (Fig. 2B), exposure of normal cardiomyocytes to 10 nM IGF-1 increased the activities of ERK1/2 (38 ± 7%; $P<.001$ and 93 ± 0.3%; $P<.05$), p38 MAPK (109 ± 16%; $P<.001$) and Akt (95 ± 13%; $P<.001$) as during eccentric hypertrophy. However, IGF-1 effect was blunted for ERK-1/2 and p38 MAPKs in the hypertrophied hearts; but not for Akt (158 ± 18%; $P=.02$ vs untreated shunt).

**Cross-talk Between ERK1/2 and p38 MAPKs Along with PI-3K in Normal and Hypertrophied Hearts**

Figure 2A shows that in the normal hearts, specific inhibition of PI-3K by LY294002 did not affect the activation level on any of the MAPKs. In the presence of the inhibitor, IGF-1 effect on ERK1/2 was abrogated but not on p38 MAPK. This may imply that IGF-1 activated p38 MAPK through a parallel pathway to PI-3K/Akt. On the other hand, PI-3K inhibition in the hypertrophied hearts reduced the basal activity of ERK1/2. However, similar to the normal hearts, inhibition of PI-3K blocked the IGF-1 effect on ERK1/2 but not on p38 MAPKs.

Figure 2B shows that in the normal hearts, specific inhibition of ERK1/2 by PD98059 did not affect the basal activation level of Akt but significantly increased that of p38 MAPK. In addition, IGF-1 effects on Akt and p38 MAPK were not altered by ERK1/2 inhibition. Conversely, in the hypertrophied hearts, ERK1/2 inhibition did not alter the basal activities of either kinase; thus the IGF-1 activation of Akt and p38 MAPK were not affected by such inhibition.

Figure 3B shows that specific inhibition of p38 MAPK by SB203580 in
the normal hearts improved the basal activity of ERK1/2 but not that of Akt. Interestingly, P38 MAPK inhibition did not affect IGF-1 activation of Akt, but blunted that of ERK1/2. In comparison, p38 MAPK inhibition in the hypertrophied hearts only enhanced ERK-2 basal activity. Such inhibition did not affect IGF-1 on Akt but prevented ERK1/2 activation.

Functional Correlate for the Combined Effects of MAPKs and Akt on Normal and Hypertrophied Cardiomyocytes

Angiotensin II (ANG II) inotropic effects are known to be mainly mediated through MAPKs and not PI-3K. Therefore, the combined effects of ANG II and IGF-1 on the cardiomyocytes would reflect, to a certain extent, the translational outcome of the cross-talk found between both pathways. Figure 3 demonstrates that exposure of normal cardiomyocytes to ANG (10 nM) with IGF-1 (10 nM) reduces the functional density of the outward rectifier potassium channel (\(I_K\)) to a greater level than in the hypertrophied ones. Interestingly, this combined effect of ANG II and IGF-1 was not altered by PI-3K inhibition in the normal cardiomyocytes, but was eliminated in the hypertrophied ones.

**DISCUSSION**

Our study shows the existence of a cross-talk between intracellular MAPKs and PI-3 kinase pathways that can be translated into a functional significance, such as the activity of potassium channels, responsible for cardiac repolarization and relaxation. It has been shown that IGF-1 effects are mainly mediated by PI-3 kinase activation.\(^1\)\(^,\)\(^2\)\(^,\)\(^13\)

We have previously confirmed this finding\(^5\) in addition to demonstrating a differential regulation of MAPKs and PI-3 kinase during the development of eccentric cardiac hypertrophy. Basically, the IGF-1 effect becomes transduced mainly via PI-3 kinase pathway. This corroborates with the reversal of the combined ANG II and IGF-1 effects by PI-3 kinase inhibition in the hypertrophied myocytes but not in the normal ones. Similar effects were observed using IGF-1 alone (data not shown).

Our data demonstrate that PI-3K has a positive influence on IGF-1-stimulated activation of ERK1/2 but not p38 MAPK in both normal and hypertrophied hearts. In addition, IGF-1-dependent ERK1/2 activation is mediated by both PI-3K and p38 MAPKs in parallel. Therefore, both kinases seem to be upstream regulator for ERK1/2 activity. Furthermore, there is a mutual negative auto-regulatory cross-talk between basal p38 MAPK and ERK1/2 activities. Such mechanism seems to be dysfunctional during the development of cardiac hypertrophy. Thus, we can
speculate that IGF-1 effects on ERK1/2 and p38 MAPK activities would be regulated to a greater extent by PI-3K during cardiac hypertrophy. This is in agreement with previous reports from our group and others that PI-3K plays a central role in mediating the hypertrophic and inotropic effects of IGF-1. Thus, the combined activation of MAPKs and PI-3K would be alleviated by PI-3K inhibition alone in the hypertrophied cardiomyocytes, but not in the normal ones. This is in accordance with our present findings on I$_K$ regulation by MAPKs and PI-3K in normal and hypertrophied cardiomyocytes.

These exciting cross-talk changes during cardiac hypertrophy are of great significance regarding the elucidation of the weaker prognostic in African Americans suffering from cardiac hypertrophy as compared to other ethnic groups. This is particularly important, since the circulating levels of free IGF-1 in African American has been reported to be more elevated than Caucasians, and more so in athletes among them.

REFERENCES